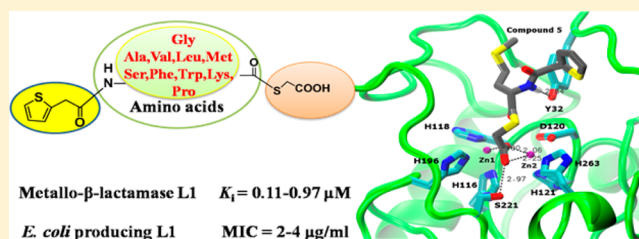


Amino Acid Thioester Derivatives: A Highly Promising Scaffold for the Development of Metallo- β -lactamase L1 InhibitorsXiao-Long Liu,^{†,||} Ying Shi,^{†,||} Joon S. Kang,[‡] Peter Oelschlaeger,[§] and Ke-Wu Yang^{*,†}[†]Key Laboratory of Synthetic and Natural Functional Molecule Chemistry of Ministry of Education, College of Chemistry and Materials Science, Northwest University, Xi'an 710127, P. R. China[‡]Department of Biological Sciences, California State Polytechnic University, Pomona, 3801 West Temple Avenue, Pomona, California 91768, United States[§]Department of Pharmaceutical Sciences, College of Pharmacy, Western University of Health Sciences, 309 East Second Street, Pomona, California 91766, United States

Supporting Information

ABSTRACT: In light of the biomedical significance of metallo- β -lactamases (M β Ls), ten new mercaptoacetic acid thioester amino acid derivatives were synthesized and characterized. Biological activity assays indicated that all these synthesized compounds are very potent inhibitors of L1, exhibiting an IC₅₀ value range of 0.018–2.9 μ M and a K_i value range of 0.11–0.95 μ M using cefazolin as substrate. Partial thioesters also showed effective inhibitory activities against NDM-1 and ImiS with an IC₅₀ value range of 12–96 and 3.6–65 μ M, respectively. Also, all these thioesters increased susceptibility of *E. coli* cells expressing L1 to cefazolin, indicated by a 2–4-fold reduction in MIC of the antibiotic. Docking studies revealed potential binding modes of the two most potent L1 inhibitors to the active site in which the carboxylate group interacts with both Zn(II) ions and Ser221. This work introduces a highly promising scaffold for the development of metallo- β -lactamase L1 inhibitors.

KEYWORDS: Antibiotic resistance, metallo- β -lactamase, subclass B3, L1, inhibitor, mercaptoacetic acid thioester



β -Lactam antibiotics, which include penicillins, cephalosporins, and carbapenems, remain the most important and frequently used antimicrobial agents, constituting more than 50% of the antibiotics prescribed worldwide.¹ However, the overuse of antibiotics in the clinical setting as well as animal production has resulted in resistance² conferred, among other resistance mechanisms, by the production of β -lactamases. More than a thousand β -lactamases have been isolated, and these enzymes have been categorized into classes A to D, depending on their amino acid sequence homologies.¹ Enzymes from classes A, C, and D comprise the serine β -lactamases. They use a catalytic mechanism that is characterized by the nucleophilic attack of an active-site serine on the β -lactam carbonyl, ultimately causing cleavage of the β -lactam ring. Class B enzymes, or metallo- β -lactamases (M β Ls), require either one or two Zn(II) ions per enzyme molecule for full catalytic activity, and these enzymes inactivate all clinically important β -lactams except aztreonam. According to amino acid sequence homologies M β Ls are divided into subclasses B1–B3, based on amino acid sequence homologies.³ M β Ls from subclasses B1 and B3 can inactivate nearly all β -lactam antibiotics. Contrarily, B2 enzymes possess a narrow substrate preference for carbapenems. There are no known clinical inhibitors of the M β Ls to date.¹

The emergence of antibiotic resistance is a global public health problem. Given the enormous clinical importance of M β Ls, there have been many attempts to identify novel M β L

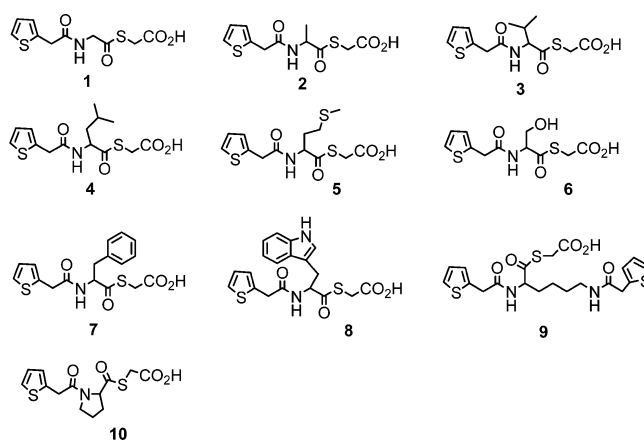


Figure 1. Structures of the synthesized mercaptoacetic acid thioesters.

inhibitors.^{4,5} Toney et al. reported biphenyl tetrazoles and succinic acids to be effective inhibitors of M β L IMP-1.⁶ Mercaptocarboxylate,⁷ 3-substituted phthalic acid derivatives,⁸ and *N*-arylsulfonyl hydrazones⁹ also displayed inhibitory

Received: February 15, 2015

Accepted: April 23, 2015

Published: April 23, 2015

Scheme 1. Synthetic Route of Mercaptoacetic Acid Thioesters

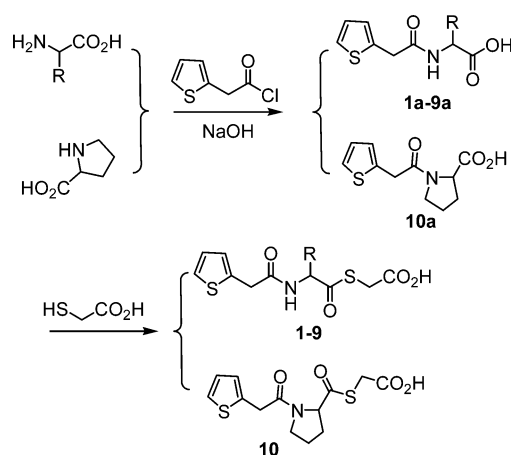


Table 1. Inhibitory Activity of Mercaptoacetic Acid Thioesters against *Mβ*Ls^a

inhibitor	IC ₅₀ (μM)			
	NDM-1 ^a	CcrA ^a	ImiS ^b	L1 ^a
1	28.7		61.5	0.84
2	95.7		46.3	0.91
3	12.4	38.5	18.9	0.45
4	28.5		3.6	0.069
5		123	13.3	0.018
6			60.1	1.49
7	36.3		65.4	2.90
8				0.29
9				1.08
10				0.45

^aBlank spaces represent no inhibition. The substrate used was cefazolin (a) and imipenem (b).

Table 2. Inhibitory Activity of Mercaptoacetic Acid Thioesters against Metallo-β-lactamase L1^a

inhibitor	K _i (μM)	inhibitor	K _i (μM)
1	0.28 ± 0.07	6	0.17 ± 0.02
2	0.41 ± 0.09	7	0.95 ± 0.09
3	0.28 ± 0.02	8	0.17 ± 0.03
4	0.21 ± 0.02	9	0.19 ± 0.02
5	0.11 ± 0.03	10	0.87 ± 0.09

^aK_i values were determined using cefazolin as substrate and L1 as the enzyme in 50 mM Tris, at 25 °C. The inhibitor concentrations were ranged from 0–2.5 μM.

Table 3. Antibacterial Activities (MIC, μg/mL) of the Mercaptoacetic Acid Thioester Inhibitors at a Final Concentration of 16 μg/mL^a

tested strains	inhibitor black	1	2	3	4	5	6	7	8	9	10
<i>E. coli</i> ^a	64	64	>64	>64	64	>64	64	64	64	64	64
<i>E. coli</i> ^b	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5
<i>E. coli</i> -NDM-1 ^a	8	4	4	4	4	8	8	8	8	4	8
<i>E. coli</i> -CcrA ^a	4	4	4	4	4	4	4	4	4	4	4
<i>E. coli</i> -ImiS ^b	2	1	1	1	1	1	1	1	2	2	2
<i>E. coli</i> -L1 ^a	8	4	2	4	2	2	4	4	2	4	2

^aThe antibiotics used were cefazolin (a) and imipenem (b).

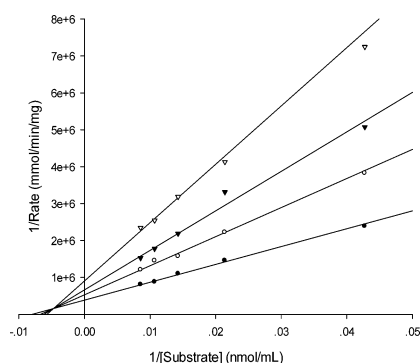


Figure 2. Lineweaver–Burk plot of L1-catalyzed hydrolysis in the absence and presence of compound 5. The substrate used was cefazolin. Inhibitor 5 concentrations were 0 μM (●), 0.625 μM (○), 1.25 μM (▼), and 2.50 μM (▽).

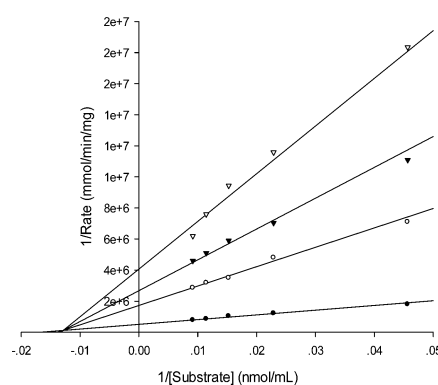


Figure 3. Lineweaver–Burk plot of L1-catalyzed hydrolysis in the absence and presence of compound 8. The substrate used was cefazolin. Inhibitor 8 concentrations were 0 μM (●), 0.625 μM (○), 1.25 μM (▼), and 2.50 μM (▽).

activities on IMP-1. A 2-picolinic acid based inhibitor was found to inhibit the B2 subclass CphA.¹⁰ Also some broad-spectrum inhibitors of the *Mβ*Ls have been reported, including mercaptophosphonate compounds, thiomandelic acid, and thiols¹¹

*Mβ*Ls inhibitors containing both sulfur and carboxylate capable of metal ligation are the most potent inhibitors.^{12–14} Payne et al. have reported several mercaptoacetic acid thioesters as irreversible inhibitors of β-lactamase BcII. Mercaptoacetic acid is delivered to the metal-coordinating cysteine as a result of the thioester conversion by the enzyme.¹⁵ However, isothermal titration calorimetry (ITC) indicated that inhibition was not accomplished through chelation of the Zn(II) ions because the presence of a phenyl substituent retarded the cleavage of the thioester bond.¹⁶ MALDI-TOF

spectrometry showed that two molecules of mercaptoacetate covalently bind to L1, a subclass B3 enzyme responsible for penicillin resistance,¹⁷ when the enzyme is incubated with benzylacetyl-D-alanylthioacetic acid. The activities of the modified enzyme and native L1 are the same.¹⁸ The structure–activity relationships (SAR) of the thioester inhibitors of the M β L IMP-1 showed that introducing more hydrophobic moieties on the C-terminus greatly enhanced their activity.¹⁹ In order to further probe the SAR of thioester inhibitors against L1 and design an effective scaffold for the development of metallo- β -lactamase L1 inhibitors, in this work, we synthesized a series of 2-thiopheneacetyl mercaptoacetic acid thioesters with various N-terminal substituents and studied their biological activities.

Ten 2-thiopheneacetyl mercaptoacetic acid thioesters (Figure 1) were synthesized by a previously reported method.^{18,20} The synthetic route is shown in Scheme 1. 2-thiopheneacetyl chloride reacted with amino acids under basic conditions to give the corresponding amides **1a–10a**. In the presence of triethylamine, the amides further reacted with mercaptoacetic acid to offer the target products 2-thiopheneacetyl mercaptoacetic acid thioesters **1–10**. These synthesized mercaptoacetic acid thioesters were characterized by ¹H and ¹³C NMR and confirmed by MS (see Supporting Information). These compounds basically represent N- and C-terminally modified derivatives of the natural amino acids glycine (**1**), alanine (**2**), valine (**3**), leucine (**4**), methionine (**5**), serine (**6**), phenylalanine (**7**), tryptophan (**8**), lysine (**9**), and proline (**10**). In **9**, two amino groups and carboxyl group of lysine are modified.

To test whether these mercaptoacetic acid thioesters were specific inhibitors of L1 or potentially broad-spectrum inhibitors of all M β Ls, M β Ls from the three subclasses (CcrA from B1a, NDM-1 from B1b, ImiS from B2, and L1 from B3) were overexpressed in *Escherichia coli* and purified as reported previously.^{4,21–23} Steady-state kinetic studies and inhibition experiments of the mercaptoacetic acid thioesters as inhibitors against M β Ls were conducted on an Agilent UV8453 spectrometer. Hydrolysis of substrate (cefazolin for CcrA, NDM-1, and L1; imipenem for ImiS) was monitored at 262 and 300 nm, respectively. The initial reaction rates were determined in the absence or presence of inhibitors in triplicate, and the average values were recorded.

The inhibitor concentrations causing 50% decrease of enzyme activity (IC₅₀) were determined and are presented in Table 1. It is clear to be observed that all mercaptoacetic acid thioesters tested effectively inhibit L1 with an IC₅₀ value range of 0.018–2.9 μ M, and **5** showed the lowest IC₅₀ value of 18 nM, revealing that mercaptoacetic acid thioesters specifically target L1. The thioesters **1–7** inhibit ImiS with an IC₅₀ value range of 4–65 μ M, while **8–10** had no inhibitory activity against ImiS. Also, apart from thioesters **3** and **5** inhibiting CcrA with IC₅₀ values of 39 and 123 μ M, respectively, the other compounds had no activity against this enzyme. It is worth noting that the mercaptoacetic acid thioesters **1–4** and **7** significantly inhibit NDM-1 with an IC₅₀ value range of 12–96 μ M, as pathogens expressing NDM-1 have been called superbugs.²⁴

Given the specificity of these mercaptoacetic acid thioesters targeting L1, in order to further identify the inhibition model of the inhibitors, we determined inhibition constants K_i of the thioesters against L1. The data listed in Table 2 show that these compounds are very potent inhibitors with a K_i value range of 0.11–0.95 μ M. The Lineweaver–Burk plots with the thioester inhibitors indicated that **1–10** are mixed inhibitors of L1.

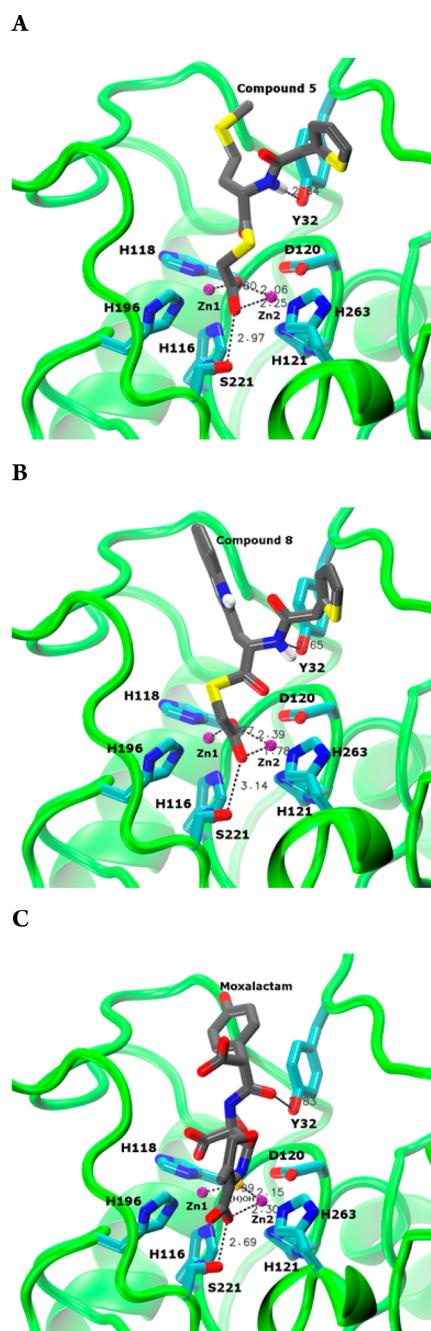


Figure 4. Lowest-energy docking conformations of compounds **5** (A) and **8** (B) docked into the active site of L1 (backbone shown as green cartoon) and hydrolyzed moxalactam cocrystallized with L1²⁶ (C) rendered as sticks (H, white; C, gray; N, blue; O, red; S, yellow). Zn(II) ions are shown as magenta spheres. The side chains of Zn(II) ligands as well as Tyr32 and Ser221 are rendered as sticks (C, cyan; N, blue; O, red). The water/hydroxide molecule (H)OH in panel C is shown as a yellow sphere. Characteristic short distances between the compounds and Zn(II) ions, (H)OH, Tyr32, and Ser221 are indicated by dashed lines.

Exemplary, inhibitors **5** and **8** are presented to show their inhibition modes against L1 in Figure 2 and 3.

The minimum inhibitory concentrations (MICs) of antibiotics in the absence or presence of compounds **1–10** were determined as previously reported method.²⁵ Four strains of *E. coli* BL21(DE3) containing plasmids pMSZ02-CcrA, pET26b-NDM-1, pET26b-ImiS, and pET26b-L1, as well as

an *E. coli* control without plasmid, were used to assess these inhibitors (Table 3). MIC data indicate that all mercaptoacetic acid thioester inhibitors resulted in 2–4-fold MIC reduction of cefazolin for *E. coli* BL21(DE3) containing the L1-encoding plasmid (*E. coli*-L1), while the MICs for the other strains were not affected significantly (no more than one dilution factor).

In order to elucidate possible binding modes of the most potent L1 inhibitors, **5** and **8**, these compounds were docked into the active site of the L1 crystal structure²⁶ using the same procedure as reported previously.²⁷ The top ranked conformations shown in Figure 4A,B exhibit common features. One of the carboxylate oxygens bridges the two Zn(II) ions, while the other oxygen coordinates Zn2 and hydrogen bonds with Ser221. Thus, one oxygen assumes the role of the bridging water/hydroxide, while the other one assumes the role of the C4 carboxylate of hydrolyzed moxalactam in the crystal structure²⁶ (Figure 4C). The hydrophobic portions of the molecules (the 2-thiopheneacetyl amido group and the amino acid side chains) are located where the C7 substituent of moxalactam is located in the crystal structure. The amide nitrogens form hydrogen bonds with the Tyr32 side chain, which is formed between the carbonyl of the C7 substituent and Tyr32 in the crystal structure. The fact that compound **6** (serine derivative) also exhibits a low K_i value (0.17 μM), while the other compounds have higher values, implies that a hydrophilic component (polar sulfur in **5**, hydroxyl group in **6**, and indole nitrogen in **8**) is favorable, possibly due to interaction with bulk solvent. Indeed, in the crystal structure the carboxylate of the moxalactam C7 substituent is in this location pointing toward the solvent. While also endowed with a hydrophilic amide group, compound **9** may simply be too big to be accommodated as efficiently in the L1 active site. Because of the combination of these specific interactions with L1, these compounds can be interpreted as analogues of a β -lactam hydrolysis product. We hope to validate this model in the near future with a crystal structure.

In summary, ten novel mercaptoacetic acid thioesters based on natural amino acids were synthesized and characterized. Biological activity assays indicated that these compounds are very potent mixed inhibitors of M β L L1 and inhibit growth of *E. coli* cells expressing L1 in the presence of cefazolin. Docking studies suggest that the carboxylate group interacts with the two Zn(II) ions and Ser221 in a similar fashion to a moxalactam hydrolysis product cocrystallized with L1. This work provides a highly promising scaffold for the development of metallo- β -lactamase L1 inhibitors.

■ ASSOCIATED CONTENT

Supporting Information

Detailed synthesis procedure, NMR and ESI mass data for target compounds; methods for enzyme overexpression and purification; inhibition assay; determination of MIC and K_i ; and docking study. These materials are available free of charge via the Internet at The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acsmchemlett.5b00098.

■ AUTHOR INFORMATION

Corresponding Author

*Tel/Fax: +8629-8153-5035. E-mail: kwyang@nwu.edu.cn.

Author Contributions

^{||}These authors contributed equally to this work.

Author Contributions

The manuscript was written through contributions of all authors.

Funding

This work was supported by grants 21272186 and 81361138018 (to K.W.Y.) from the National Natural Science Foundation of China and key grant 2014KW23-03 (to K.W.Y.) for international cooperation of Shaanxi province.

Notes

The authors declare no competing financial interest.

■ REFERENCES

- (1) Drawz, S. M.; Bonomo, R. A. Three decades of β -lactamase inhibitors. *Clin. Microbiol. Rev.* **2010**, *23*, 160–201.
- (2) Fisher, J. F.; Meroueh, S. O.; Mobashery, S. Bacterial resistance to β -lactam antibiotics: Compelling opportunism, compelling opportunity. *Chem. Rev.* **2005**, *105*, 395–424.
- (3) Bebrone, C. Metallo- β -lactamases (classification, activity, genetic organization, structure, zinc coordination) and their superfamily. *Biochem. Pharmacol.* **2007**, *74*, 1686–701.
- (4) Yang, H.; Aitha, M.; Hetrick, A. M.; Richmond, T. K.; Tierney, D. L.; Crowder, M. W. Mechanistic and Spectroscopic Studies of Metallo- β -lactamase NDM-1. *Biochemistry* **2012**, *51*, 3839–47.
- (5) Bush, K. The ABCD's of β -lactamase nomenclature. *J. Infect. Chemother.* **2013**, *19*, 549–59.
- (6) Toney, J. H.; Hammond, G. G.; Fitzgerald, P. M.; Sharma, N.; Balkovec, J. M.; Rouen, G. P.; Olson, S. H.; Hammond, M. L.; Greenlee, M. L.; Gao, Y. D. Succinic acids as potent inhibitors of plasmid-borne IMP-1 metallo- β -lactamase. *J. Biol. Chem.* **2001**, *276*, 31913–8.
- (7) Concha, N. O.; Janson, C. A.; Rowling, P.; Pearson, S.; Cheever, C. A.; Clarke, B. P.; Lewis, C.; Galleni, M.; Frere, J. M.; Payne, D. J.; Bateson, J. H.; Abdel-Meguid, S. S. Crystal structure of the IMP-1 metallo- β -lactamase from *Pseudomonas aeruginosa* and its complex with a mercaptocarboxylate inhibitor: Binding determinants of a potent broad-spectrum inhibitor. *Biochemistry* **2000**, *39*, 4288–98.
- (8) Hiraiwa, Y.; Morinaka, A.; Fukushima, T.; Kudo, T. Metallo- β -lactamase inhibitory activity of phthalic acid derivatives. *Bioorg. Med. Chem. Lett.* **2009**, *19*, 5162–5.
- (9) Siemann, S.; Evanoff, D. P.; Marrone, L.; Clarke, A. J.; Viswanatha, T.; Dmitrienko, G. I. N-Arylsulfonyl hydrazones as inhibitors of IMP-1 metallo- β -lactamase. *Antimicrob. Agents Chemother.* **2002**, *46*, 2450–7.
- (10) Horsfall, L. E.; Garau, G.; Lienard, B. M.; Dideberg, O.; Schofield, C. J.; Frere, J. M.; Galleni, M. Competitive inhibitors of the CphA metallo- β -lactamase from *Aeromonas hydrophila*. *Antimicrob. Agents Chemother.* **2007**, *51*, 2136–42.
- (11) Lienard, B. M.; Garau, G.; Horsfall, L.; Karsisiotis, A. I.; Damblon, C.; Lassaux, P.; Papamichael, C.; Roberts, G. C.; Galleni, M.; Dideberg, O.; Frere, J. M.; Schofield, C. J. Structural basis for the broad-spectrum inhibition of metallo- β -lactamases by thiols. *Org. Biomol. Chem.* **2008**, *6*, 2282–94.
- (12) Garcia-Saez, I.; Hopkins, J.; Papamichael, C.; Franceschini, N.; Amicosante, G.; Rossolini, G. M.; Galleni, M.; Frère, J. M.; Dideberg, O. J. Structure of *Chryseobacterium meningosepticum* zinc β -lactamase in complex with the inhibitor, D-captopril. *Biol. Chem.* **2003**, *278*, 23868–73.
- (13) Wachino, J.; Yamaguchi, Y.; Mori, S.; Kurosaki, H.; Arakawa, Y.; Shibayama, K. Structural insights into the subclass B3 metallo- β -lactamase SMB-1 and the mode of inhibition by the common metallo- β -lactamase inhibitor mercaptoacetate. *Antimicrob. Agents Chemother.* **2013**, *57*, 101–9.
- (14) Faridooon; Islam, N. U. An update on the status of potent inhibitors of metallo- β -lactamases. *Sci. Pharm.* **2013**, *81*, 309–27.
- (15) Payne, D. J.; Bateson, J. H.; Gasson, B. C.; Proctor, D.; Khushi, T.; Farmer, T. H.; Tolson, D. A.; Bell, D.; Skett, P. W.; Marshall, A. C.; Reid, R.; Ghosez, L.; Combret, Y.; Marchand-Brynaert, J. Inhibition of

metallo- β -lactamases by a series of mercaptoacetic acid thiol ester derivatives. *Antimicrob. Agents Chemother.* **1997**, *41*, 135–40.

(16) Payne, D. J.; Bateson, J. H.; Gasson, B. C.; Khushi, T.; Proctor, D.; Pearson, S. C.; Reid, R. Inhibition of metallo- β -lactamases by a series of thiol ester derivatives of mercaptophenylacetic acid. *FEMS Microbiol. Lett.* **1997**, *157*, 171–5.

(17) Yang, K. W.; Crowder, M. W. Inhibition studies on the metallo- β -lactamase L1 from *Stenotrophomonas maltophilia*. *Arch. Biochem. Biophys.* **1999**, *368*, 1–6.

(18) Feng, L.; Yang, K. W.; Zhou, L. S.; Xiao, J. M.; Yang, X.; Zhai, L.; Zhang, Y. J.; Crowder, M. W. N-Heterocyclic dicarboxylic acids: broad-spectrum inhibitors of metallo- β -lactamases with co-antibacterial effect against antibiotic-resistant bacteria. *Bioorg. Med. Chem. Lett.* **2012**, *22*, 5185–9.

(19) Greenlee, M. L.; Laub, J. B.; Balkovec, J. M.; Hammond, M. L.; Hammond, G. G.; Hammond, M. L.; Pompliano, D. L.; Epstein-Toney, J. H. Synthesis and SAR of thioester and thiol inhibitors of IMP-1 metallo- β -lactamase. *Bioorg. Med. Chem. Lett.* **1999**, *9*, 2549–54.

(20) Banerjee, P. S.; Zuniga, E. S.; Ojima, I.; Carrico, I. S. Targeted and armed oncolytic adenovirus via chemoselective modification. *Bioorg. Med. Chem. Lett.* **2011**, *21*, 4985–8.

(21) Wang, Z.; Benkovic, S. J. Purification, characterization, and kinetic studies of a soluble *Bacteroides fragilis* metallo- β -lactamase that provides multiple antibiotic resistance. *J. Biol. Chem.* **1998**, *273*, 22402–8.

(22) Crawford, P. A.; Sharma, N.; Chandrasekar, S.; Sigdel, T.; Walsh, T. R.; Spencer, J.; Crowder, M. W. Over-expression, purification, and characterization of metallo- β -lactamase ImiS from *Aeromonas Veronii* bv. *sobria*. *Protein Expr. Purif.* **2004**, *36*, 272–9.

(23) Crowder, M. W.; Walsh, T. R.; Banovic, L.; Pettit, M.; Spencer, J. Overexpression, purification, and characterization of the cloned metallo- β -lactamase L1 from *Stenotrophomonas maltophilia*. *Antimicrob. Agents Chemother.* **1998**, *42*, 921–6.

(24) Ma, J.; McLeod, S.; MacCormack, K.; Sriram, S.; Gao, N.; L. Breeze, A. L.; Hu, J. Real-time monitoring of New Delhi metallo- β -lactamase activity in living bacterial cells by ^1H NMR spectroscopy. *Angew. Chem., Int. Ed.* **2014**, *53*, 2130–3.

(25) Xiao, J. M.; Feng, L.; Zhou, L. S.; Gao, H. Z.; Zhang, Y. L.; Yang, K. W. Novel fluorescent cephalosporins: synthesis, antimicrobial activity and photodynamic inactivation of antibiotic resistant bacteria. *Eur. J. Med. Chem.* **2013**, *59*, 150–9.

(26) Spencer, J.; Read, J.; Sessions, R. B.; Howell, S.; Blackburn, G. M.; Gamblin, S. J. Antibiotic recognition by binuclear metallo- β -lactamases revealed by X-ray crystallography. *J. Am. Chem. Soc.* **2005**, *127*, 14439–44.

(27) Zhang, Y. L.; Yang, K. W.; Zhou, Y. J.; LaCuran, A. E.; Oelschlaeger, P.; Crowder, M. W. Diaryl-substituted azolythioacetamides: inhibitor discovery of New Delhi metallo- β -lactamase-1 (NDM-1). *ChemMedChem* **2014**, *9*, 2445–8.